Genomic analysis of diversity, population structure, virulence, and antimicrobial resistance in *Klebsiella pneumoniae*, an urgent threat to public health

Kathryn E. Holt^{a,b,1}, Heiman Wertheim^{c,d}, Ruth N. Zadoks^{e,f}, Stephen Baker^g, Chris A. Whitehouse^h, David Dance^{d,i}, Adam Jenney^{b,j}, Thomas R. Connor^{k,I}, Li Yang Hsu^m, Juliëtte Severinⁿ, Sylvain Brisse^o, Hanwei Cao^{b,p}, Jonathan Wilksch^{b,p}, Claire Gorrie^{a,b,p}, Mark B. Schultz^a, David J. Edwards^a, Kinh Van Nguyen^q, Trung Vu Nguyen^q, Trinh Tuyet Dao^q, Martijn Mensink^e, Vien Le Minh^{g,r}, Nguyen Thi Khanh Nhu^{g,s}, Constance Schultsz^{g,t}, Kuntaman Kuntaman^u, Paul N. Newton^{d,i}, Catrin E. Moore^{d,i}, Richard A. Strugnell^{b,p}, and Nicholas R. Thomson^{k,v,1}

PNAS

Journal Club 14.09.2016 Maido Remm

Eesmärk

- *Klebsiella pneumoniae* is rapidly becoming untreatable using last-line antibiotics. It is especially problematic in hospitals, where it causes a range of acute infections.
- To approach controlling such a bacterium, we first must define what it is and how it varies genetically.

Mida tehti

- Sekveneeriti 288 erinevat *K. pneumoniae* isolaati (tüve), mis pärinesid 4 kontinendilt. Lisaks 40 tüve NCBI andmebaasist
- Proove võeti nii haigetelt kui tervetelt inimestelt, lisaks ka lehmadelt ja keskkonnast.
- Analüüsid:
 - Populatsiooni fülogeneetiline struktuur
 - Geenide arv genoomides
 - Geenide ja virulentsuse seoste leidmine

DNA eraldamine ja sekveneerimine:

- Genomic DNA was extracted at each site using either phenol-chloroform extraction, QIAmp (Qiagen), High Pure or MagNA Pure (Roche).
- Index-tagged paired end Illumina sequencing libraries were prepared using one of 12 unique indexing tags (9), combined into pools of uniquely tagged libraries and sequenced on the Illumina Genome Analyzer GAII at the Wellcome Trust Sanger Institute to generate tagged
 76 bp paired-end reads.





Kas need 3 liiki ristuvad omavahel?



Density of SNPs per kilobase along the genome of Vietnamese human gut carriage isolate D022 compared to reference genomes from KpI (**a**) and KpII (**b**). The 735 kbp region of proposed homologous recombination between D022 and a KpII-like genome is shown in purple and bounded by dashed lines.

=> There are no obvious mechanistic barriers to homologous recombination between KpI, KpII, and KpIII; indeed the observation of a large recombination between KpI and KpII shows that homologous recombination is possible, although the rarity of this event (1 out of >300 genomes) suggests there could be selection against such hybrids.

=> The speciation of KpII and KpIII is likely driven by long-term separation in distinct ecological niches.

Klebsiella liigid on erinevate omadustega

C	КрІ	KpII-B	KpIII	p-value (Chisq)	N (% missing)
Source type:					
Human (vs any other)	72%	94%	50%	*0.01	266 (1.8%)
Bovine (vs any other)	22%	0%	50%	*0.002	
Infection status:					
Infections (vs colonization)	71%	40%	61%	*0.03	245 (9.6%)
Noscomial	30%	56%	22%	0.06	237 (12.5%)
(vs community acquired)					
Invasive	26%	0%	22%	*0.05	166 (1.8%)
(vs non-invasive infection)					
Death (vs discharge)	26%	0%	0%	0%	81 (70%)

Klebsiella liigid on erinevate omadustega



Pan-genoom



Unique protein clustered at the ≤30% amino acid homology level using CD-HIT



Gene frequency (# K. pneumoniae genomes)

Pan-genome: Kokku 29,886 erinevat geeni
ca 5000 geeni esineb vaid <u>ühes tüves</u>
1743 geeni esineb <u>kõigis tüvedes</u>
Accessory genome: 17 700 geeni esineb >5% ja <95% tüvedes

Figure S2 - Accessory genes in K. pneumoniae



Maximum likelihood tree of 328 *K. pneumoniae* isolates (left), with heatmap indicating presence (black) or absence (white) of 7,771 accessory genes that are present in 5%-95% of genomes (right).

Geenide jaotumine sugupuu järgi

Virulentsusgeenide jaotumine tüvedes





Kogu info ühes tabelis:



Kas virulentsusgeenide suhteline sagedus on invasiivse fenotüübiga tüvedes erinev?



Invasiivne on tüvi, mis on isoleeritud kudedest, mis on tavaliselt steriilsed

PGWAS

 To extend these observations, we performed a PGWAS, screening each gene in the KpI pangenome for association with infection in humans. The strongest associations, reaching pan-genome-wide significance after correcting for multiple testing, were the rmpA/2 and siderophore genes and five additional predicted iron-metabolism genes (OR 8–10; 95% CI 3–35), also present on the virulence plasmid pK2044.

Gene screen for invasive infection



Each gene was screened for association with (**a**) infection (vs. carriage) and (**b**) invasive infection (vs. non-invasive infection or carriage), amongst human KpI isolates, using Fisher's exact test. Black bars indicate the distribution of the resulting p-values (note P-values on the x-axis are shown on the natural log scale). P-values for known virulence gene clusters are indicated with labelled vertical bars, coloured as in Figure 4; solid lines indicate significant associations ($P \le 0.05$, i.e. $-\log(P) \ge 3$); dashed lines indicate non-significant associations ($P \ge 0.05$, i.e. $-\log(P) \ge 3$); dashed lines indicate non-significant associations ($P \ge 0.05$, i.e. $-\log(P) \ge 3$); P-values for novel virulence genes are indicated with black lines: 1, pK2044_00025 (FepB domain); 2, pK2044_00325 (CobW domain); 3, pK2044_00355 (Fur domain); 4, pK2044_00335 (FeoB domain); 5, pK2044_00030 (FepC domain).

Fülogeneetilise tausta võimaliku mõju taandamiseks kasutati logistilist regressiooni

- **Yersiniabactin**, whose synthesis is encoded by the ybt, irp1, irp2, and fyuA genes, was the most prevalent virulence-associated locus, present in one third of the KpI human isolates.
- Despite this high prevalence it was a strong predictor of infection vs. carriage in humans, with an OR of 7.4 [95% confidence interval (CI), 2.2–40; P = 0.0001; Fisher's exact test] and a positive predictive value of 95%.
- This effect was not dependent on chromosomal background, because yersiniabactin was significantly associated with infection in a logistic regression model that included phylogenetic lineage (OR 1.3; P = 0.003).

Pangenome analysis

- Illumina reads were assembled using the *de novo* short read assembler Velvet and Velvet Optimiser (19). Publicly available finished or draft genomes were also included. Contigs less than 100 bp in size were excluded from further analysis. We then used an iterative mapping approach as described previously (20), to generate a pangenome representing the nonredundant set of all >5% divergent sequences of length ≥100 bp among the 328 *K. pneumoniae* (using the nucmer algorithm in MUMmer).
- Open reading frames (ORFs) were identified and annotated using Prokka (23) with primary reference to NTUH-K2044 protein sequences. **A total of 84,175 ORFs were identified**, which were unique at the 5% DNA homology level. All *K. pneumoniae* read sets were aligned to this pangenome sequence.
- The ORFs were translated into protein using EMBOSS and clustered at the ≤30% amino acid homology level using CD-HIT (24), resulting in 29,886 protein/gene clusters.
- Alternative clustering at ≤10%, ≤20% or ≤40% homology resulted in 46718, 37609 or 29779 clusters, respectively; hence 30% was taken as the point of inflection.
- These data were then processed to generate a binary gene content matrix in which the
 presence of a gene is defined as >90% coverage of at least one ORF belonging to the
 corresponding protein cluster. Hence genomes that encode alleles with ≥30% amino acid
 homology across the length of the sequence are considered to encode the same functional
 gene in our analysis.