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Sequencing platform bias

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Media Registration GENOME INFORMATICS October 30 - November 2, 2013 Abstract Deadline: September 3, 2013

Organizers: Jennifer Harrow, Wellcome Trust Sanger Institute, UK Michael Schatz, Cold Spring Harbor Laboratory James Taylor, Emory University



A note on high coverage NGS SNP reproducibility

KEYNOTE SPEAKER I

Dr. Michael Snyder

Stanford University School of Medicine

GENOME INFORMATICS

"Personal Omics Profiling: Monitoring Genomes, Transcriptomes and other Omes During Healthy and Disease States"

Dr. Michael Snyder

Stanford University School of Medicine

- Same sample sequenced twice with same methods
- Illumina, 60x coverage
- Run 1: 3.948 M SNPs called
- Run 2: 3.925 M SNPs called
- 6% of total 4 M unique SNPs don't overlap due to no-calls
- Of the overlapping SNPs, 0.05% are discordant

GENOME INFORMATICS

Dr. Michael Snyder





nature biotechnology

Performance comparison of whole-genome sequencing platforms

Hugo Y K Lam^{1,8}, Michael J Clark¹, Rui Chen¹, Rong Chen^{2,8}, Georges Natsoulis³, Maeve O'Huallachain¹, Frederick E Dewey⁴, Lukas Habegger⁵, Euan A Ashley⁴, Mark B Gerstein^{5–7}, Atul J Butte², Hanlee P Ji³ & Michael Snyder¹

Complete Genomics vs. Illumina

Presentation emphasis on overlap and concordance of SNVs (single nucleotide variants)

- 1 individual sequenced on Illumina and CG platform
- 2 DNA sources: blood (mononuclear cells) and saliva

- Illumina HiSeq 2000: 101-bp paired-end reads
- CG: 35-bp paired-end reads
- Illumina
 - mapped with BWA
 - SNVs called with GATK
- CG mapped and called with CG pipeline



 Table 1 Whole-genome sequencing using CG and Illumina platfor

| | CG | | | |
|--------|------------|--------------|------------|--------------|
| Sample | Bases (Gb) | Coverage (×) | Bases (Gb) | Coverage (×) |
| Blood | 233.2 | 78 | 151.4 | 50 |
| Saliva | 218.6 | 73 | 307.1 | 102 |
| Tatal | 151 Q | 151 | 152 5 | 152 |

- Data from blood and saliva combined
- SNVs detected from only one source discarded
- Only few of the tissue-specific calls could be validated by independent methods



Independent validation

Omni Quad 1M Genotyping array

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Agilent SureSelect target enrichment for total 33,084 SNVs

 Validated 92.7% of concordant; 61.9% CG-specific; 64.3% Illumina-specific SNVs

Lam et al. 2012

CG specific

IL specific

CG+IL

Ca. **3.3 million** overlapping SNVs

2.7%

88.1% of total unique SNVs98.9% of the overlapping are concordant

Sensitivity **99.34**% of Illumina Omni array

9.2%

Lam et al. 2012



- ti/tv of concordant SNVs was very close to that expected, whereas the platform-specific ti/tv was much lower
- Quality scores of platform-specific SNVs were lower



- Average GC content
 - Concordant: 0.46
 - CG-specific: 0.45
 - Illumina-specific: 0.41

• Average read depths: 48, 47 and 44, respectively.

• No strong correlation of SNV detection with GC content.

- Platform-specific SNVs associated with repetitive elements such as Alu, telomere and simple repeat sequences
- Only 0.3% of the concordant SNVs in telomeres or centromeres, opposed to 4% of CG-specific and 2% of Illumina-specific



Indel calls are greatly discordant



Conclusions

- Each genome sequencing approach is generally capable of detecting most SNVs
- CG appears to be more accurate, but also slightly less sensitive
- Ilumina covers more bases and makes a higher number of overall calls, but also has more false positives

Conclusions

- Each genome sequencing approach is generally capable of detecting most SNVs
- CG appears to be more accurate, but also slightly less sensitive
- Ilumina covers more bases and makes a higher number of overall calls, but also has more false positives
- Both methods clearly call variants missed by the other technology
- Filtering for repeat regions and quality helps to reduce errors
- For maximized sensitivity and quality, sequence in parallel

Comparison of Sequencing Platforms for Single Nucleotide Variant Calls in a Human Sample

Aakrosh Ratan^{1®}, Webb Miller¹, Joseph Guillory², Jeremy Stinson², Somasekar Seshagiri², Stephan C. Schuster^{1*®}

1 Center for Comparative Genomics and Bioinformatics, Pennsylvania State University, University Park, Pennsylvania, United States of America, 2 Department of Molecular Biology, Genentech Inc., South San Francisco, California, United States of America

- 454/Roche GS FLX
- Illumina HiSeq 2000
- ABI SOLID 3 ECC
- Overlap of SNVs
- 1 DNA sample

| | 454 | Illumina | SOLID |
|---------------------------|-------------------------|--------------------------|------------------------------|
| Number of reads generated | 83,331,227 | 1,867,073,052 | 6,905,193,148 |
| Number of bases generated | 29,246,232,549 | 188,349,876,745 | 397,681,271,500 |
| Read lengths | 350 avg. single-end | 101 paired-end | 50 paired-end, 75 single-end |
| Number of reads aligned | 82,310,265 (98.77%) | 1,751,042,389 (93.79%) | 4,429,505,837 (64.15%) |
| Number of bases aligned | 28,732,501,185 (98.24%) | 168,495,777,999 (89.46%) | 224,998,686,646 (56.58%) |
| Coverage | 10.04/9.78 X | 58.89/55.06 X | 78.63/53.20 X |
| Duplicate reads | 2,211,903 | 115,528,614 | 1,216,108,795 |
| Reference bases covered | 2,781,827,482 | 2,858,458,440 | 2,850,277,778 |

Table 1. Sequencing and alignment statistics. Coverage is calculated with and without the putative PCR duplicates.

The number of aligned reads includes the duplicate reads. doi:10.1371/journal.pone.0055089.t001

- Uneven coverage between platforms
- "we sequenced the individual's DNA to read-depths that allows for variant detection in each corresponding dataset with sufficient confidence "



Removing the sex chromosomes from the analysis did not eliminate the bimodal behavior



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| | | | | Lam et al. 2012 |

Variation of coverage with GC content

Illumina(HiSeq)



SOLID



Variation of coverage with GC content



Ratan et al. 2013

Numbers of detected variants

- 454: 4,331,131
- Illumina: 4,691,363
- SOLiD: 4,145,208
- Total combined: 5,252,985
- Shared between three: 3,401,954 (64.8%)

Numbers of detected variants – 5.25 M total



Ratan et al. 2013

- "self-masking" to identify the regions in the reference genome where reads should align uniquely – LASTZ
- reference genome broken into fragments of length equal to the length of the reads
- Fragments mapped back to reference
- Considering only uniquely mappable regions
- Very useful for 454 data
- Less so for Illumina and SOLiD

Validation using Sequenom Mass Spectrometry



Reducing platform bias in next-generation sequencing.

Lauri Saag^{1,2}, Ulvi Gerst Talas¹, Mario Mitt^{1,3}, Reedik Mägi³, Simon Rasmussen⁴, Richard Villems^{1,2} and Mait Metspalu²

Poster at ASHG 2013 in Boston

Complete Genomics vs. Illumina

Concordance of base calls in potential SNP positions

Study design.

- Genomes of 7 individuals sequenced on both platforms.
- Concordance measurements presented for 4 samples with good and similar sequence statistics.
- CG: average coverage 40x, processed with CG pipeline.
- Illumina: average coverage 26x, mapping with BWA, multisample calling with SAMtools.

Study design.

- Positions selected where SNP occurred in at least one individual in a dataset of 249 humans from various populations worldwide (sequenced by CG). Sites with other types of mutations excluded.
- From this list of ca. 50 million SNP sites, we selected those for which information (either reference, variant or no-call) was available from both platforms and all samples.
- Thus, our comparison is based on the call concordance in predefined sites, and not on the overlap of variant-only data.
 If a site is discordant between platforms, it appears as private for both.

Initial filtering.

- CG: VQHIGH (PASS) filter.
- Illumina: Read depth min 10, max 100; males X, Y chromosomes min 5, max 50. Variants in repeats removed.
- Ambiguous bases (N) in compared files may be either no-calls or filtered.

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Mismapping positions filter.

- We masked the data with windows in the genome where mapping of Illumina reads often fails.
- Applicability of these blacklists to CG data was not known.

- Total 48.6 M sites, 39 M unambiguous (not N) in both platforms.
- 29.2 M sites concordant in all 4 samples, 28.8 M unambiguous.
- Averages over 4 samples, given for unambiguous discordances (percent ranges ±0.1)



private Initial filters: (9.4 M total) (1.7 M Ns)1.3 M unambig. 3.3% Mismapping filter: (6.8 M total) (0.88 M Ns) 0.91 M unambig. 3.2%

CG

CG+IL Concordant: same base, same zygocity, not N in either platforms. Initial filters: (39.2 M total) (1.5 M Ns) 37.7 M unambig. 96,7%

> Mismapping filter: (28.7 M total) (1 M Ns) 27.7 M unambig. **96,8%**

private

IL

Initial filters: (9.4 M total) (6.5 M Ns) 1.3 M unambig. **3.3%**

Mismapping filter: (6.8 M total) (5 M Ns) 0.91 M unambig. **3.2%**

- Poor-mapping windows only minimally reduce the bias.
- The bias in these positions is not greater than elsewhere, suggesting that if errors are indeed present in these positions in our Illumina data then the current version of our windows may also apply to CG.

Concordant positions filter and additional 3 samples.

- 29.2 M sites were concordant in all 4 samples.
- We used these as a whitelist to filter the sequences of other 3 samples that had lower average quality and large numbers of ambiguous sites.

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- 29.2 M sites were concordant in all 4 samples.
- We used these as a whitelist to filter the sequences of other 3 samples that had lower average quality and large numbers of ambiguous sites.
- On average, from 22.3 M unambiguous sites in both platforms 18.1 remained (19%) and the bias dropped to 1.4% from 3.4%.
- Despite lower quality, the additional 3 samples showed stable and very similar bias percentage as the other four: 3.4% after initial filtering, 3.3% with mismapping filter and 3.1% with biased positions filter.